

研究论文

环状RNA在胃癌细胞系中的差异表达

龙志¹, 谢娟², 刘艳萍¹, 张琳¹, 徐晨光¹, 罗江艳¹, 李国庆^{1,*}

¹南华大学附属第二医院消化内科; ²南华大学医学院诊断教研室, 衡阳 421001

摘要: 本文旨在探讨环状RNA (circular RNA, circRNA)在不同分化程度的胃癌细胞MGC-803、SGC-7901、NCI-N87与正常胃上皮细胞GES-1中的表达谱变化。体外培养低分化胃癌细胞MGC-803、中分化胃癌细胞SGC-7901、高分化胃癌细胞NCI-N87和正常胃上皮细胞GES-1, 利用高通量circRNA芯片技术检测三种不同分化程度胃癌细胞和正常胃上皮细胞中差异表达的circRNA, 借助生物信息学软件预测可能与circRNA相互作用的微小RNA (microRNA, miRNA), 结合文献检索初步选定可能在胃癌中具有重要意义的circRNA。结果显示, 与正常胃上皮细胞相比, 在不同分化程度胃癌细胞中均表达上调的circRNA有79条, 均表达下调的有229条。生物信息学分析和文献检索结果显示, hsa_circ_0001897与胃癌分期、转移相关, 与其结合的miR-150-5p与多种消化道肿瘤密切相关, 如结肠癌、肝癌、胆管癌; 与hsa_circ_0008106、hsa_circ_0060456存在结合位点的miR-16-5p已被证实参与胃癌的发生和发展。上述结果提示, hsa_circ_0001897、hsa_circ_0008106、hsa_circ_0060456可能通过与miRNA相互作用参与胃癌的发生和发展。

关键词: 环状RNA; 胃癌; 基因芯片

中图分类号: R735.2

Differentially expressed circular RNAs in human gastric cancer cells

LONG Zhi¹, XIE Juan², LIU Yan-Ping¹, ZHANG Li¹, XU Chen-Guang¹, LUO Jiang-Yan¹, LI Guo-Qing^{1,*}

¹Department of Gastroenterology, the Second Affiliated Hospital of University of South China; ²Teaching and Research Section of Diagnosis, School of Medicine, University of South China, Hengyang 421001, China

Abstract: The purpose of this study was to investigate the expression profile of circular RNA (circRNA) in gastric cancer cells MGC-803, SGC-7901 and NCI-N87 with different degrees of differentiation and normal gastric epithelial cells GES-1. High throughput circRNA microarray technique was used to detect the differential expression of circRNA between three kinds of differentiated gastric cancer cells and normal gastric epithelial cells. The interaction of microRNAs (miRNAs) with circRNAs was predicted by bioinformatics software, and circRNA, which might have great significance in gastric cancer, was identified by literature search. The results showed that there were 79 up-regulated circRNAs and 229 down-regulated circRNAs in gastric cancer cells with different degrees of differentiation compared with those in normal gastric epithelial cells. Through bioinformatics software analysis and literature retrieval, it was found that hsa_circ_0001897 was related to the staging and metastasis of gastric cancer, while the miR-150-5p, which combined with it, was closely related to many kinds of digestive tract tumors, such as colon cancer, liver cancer, and cholangiocarcinoma. The miR-16-5p, which has binding sites with hsa_circ_0008106 and hsa_circ_0060456, has been confirmed to be involved in the development of gastric cancer. The above results suggest that hsa_circ_0001897, hsa_circ_0008106 and hsa_circ_0060456 may be closely related to the occurrence and development of gastric cancer by interacting with miRNA.

Key words: circRNA; gastric cancer; microarray

Received 2018-03-27 Accepted 2018-05-09

This work was supported by the Second Batch of Research Projects in 2017 from Hunan Provincial Health and Family Planning Commission (No. A2017014) and the National Natural Science Foundation of China (No. 81071965).

*Corresponding author. Tel: +86-734-8899681; E-mail: ligq1970@163.com

胃癌是世界上最常见的癌症之一，我国胃癌发病例数和死亡例数分别占全球胃癌发病和死亡的42.6%和45.0%，在全球183个国家中位于发病率第5位、死亡率第6位^[1]。目前，虽然癌症基因、抑癌基因和与胃癌相关的肿瘤信号通路已经被发现和证实，但是由于我们对多基因参与肿瘤的调控步骤的认识有限，并且缺乏有效的干预手段，胃癌的诊治及预后仍不乐观。因此，寻找新的生物靶点来监测和控制胃癌的发生和发展具有重要的科学和实用价值。

环状RNA(circular RNA, circRNA)是一类由特殊的前体RNA(pre-RNA)通过反向剪接的方式产生的新型非编码RNA^[2]。研究人员已经证实circRNA能够与微小RNA(microRNA, miRNA)相互作用，从而参与结肠癌、肝癌、肺癌等肿瘤的发生和发展^[3-5]。本研究在不同分化程度胃癌细胞与正常胃上皮细胞中，利用高通量circRNA芯片技术筛选出差异表达的circRNA，借助生物信息学软件预测可能与circRNA相互作用的miRNA，为深入探讨circRNA在胃癌发生和发展中的作用机制奠定基础，同时也为胃癌的诊治提供新的思路。

1 材料和方法

1.1 主要试剂 Trizol试剂购自美国Invitrogen公司；RNase R购自美国Epicentre公司；Rneasy Mini Kit购自德国QIAGEN公司；Cy3-dCTP购自美国GE Healthcare公司；SuperScriptTM III Reverse Transcriptase购自美国Invitrogen公司；RPMI-1640培养基、无支原体胎牛血清购自美国GIBCO公司；0.25%Trypsin-EDTA胰酶购自美国Hyclone公司；DMSO购自美国Sigma公司；Arraystar circRNA芯片(8×15K)由上海康成生物公司提供。

1.2 细胞培养 正常胃上皮细胞GES-1购自中南大学湘雅医学院细胞中心，人低分化胃癌细胞株MGC-803购自上海吉玛制药技术有限公司，人中分化胃癌细胞株SGC-7901购自上海吉凯基因公司，人高分化胃癌细胞株NCI-N87购自中国科学院干细胞库，用RPMI-1640培养基(含10%胎牛血清)，在培养箱内于5%CO₂、37℃条件下培养。

1.3 样品总RNA的提取和质控 待细胞完全贴壁后，弃去培养液，按照Trizol试剂说明书，从不同分化程度的胃癌细胞株MGC-803、SGC-7901、NCI-N87和正常胃上皮细胞GES-1中提取总RNA，

应用Nanodrop-1000分光光度计分别检测各个样本在230 nm、260 nm及280 nm的吸光度值，计算RNA浓度和质量。

1.4 circRNA芯片数据采集 将上述获得的总RNA用RNase R处理以去除线性RNA分子，按照Arraystar Super RNA Labeling protocol对circRNA进行扩增并转录成荧光cRNA，用Rneasy Mini Kit对标记的cRNA进行纯化，然后放在circRNA芯片(8×15K)上杂交，于杂交炉中65℃孵育17 h，杂交反应结束后，取出芯片，首先在42℃含0.2%SDS的2×SSC的洗液中洗涤5 min，然后在室温下，将芯片放入0.2×SSC的洗液中洗涤5 min，取出芯片甩干，将洗片固定在Agilent Scanner G2505C扫描仪上，检测芯片的荧光强度。将芯片所扫描的图片导入Agilent Feature Extraction(版本11.0.1.1)软件以提取数据，然后运用R软件包对所得数据进行标准化并进一步分析，样本间差异表达的circRNA通过倍数变化(fold change, FC, FC>1.5)和P值(<0.05)进行筛选，实验重复3次。两样本间比较采用t检验，当P<0.05时，差异具有统计学意义。circRNA芯片数据的采集、以及火山图与聚类图的绘制和分析由上海康成生物公司完成。

1.5 生物信息学分析 根据差异表达FC和P值筛选circRNA分子，利用生物信息学软件Target-scan和Miranda预测与circRNA存在结合位点的miRNA。

2 结果

2.1 总RNA、cRNA的质量检测

分光光度计结果显示，胃癌细胞实验组和正常胃上皮细胞对照组RNA OD₂₆₀/OD₂₈₀均为1.8~2.0，OD₂₆₀/OD₂₃₀均>1.8，RNA总量均≥0.5 μg，cRNA总量均≥1.65 μg，Cy3标记特异活性均≥9 pmol Cy3/μg，总RNA与cRNA质量检测均符合实验要求(表1)，可用于下一步芯片杂交实验。

2.2 胃癌中差异表达的circRNA

差异表达的circRNA筛选标准为：FC>1.5，P<0.05。芯片结果显示，与正常胃上皮细胞GES-1相比，在低分化胃癌细胞MGC-803、中分化胃癌细胞SGC-7901和高分化胃癌细胞NCI-N87中均表达上调的circRNA有79条，均表达下调的有229条。表2、表3列出了表达上调和下调的前5条circRNA。

2.3 绘制火山图和聚类图分析表达谱差异

根据上述芯片差异表达谱绘制火山图和聚类图。火山图如图 1 所示，红点代表在胃癌细胞中差异表达倍数在 1.5 倍以上， P 值小于 0.05 的 circRNA。聚类图如图 2 所示，红色区域代表 circRNA 相对表达量高，绿色代表 circRNA 相对表达量低，差异表达的 circRNA 能有效区分胃癌细胞和正常胃上皮细胞。

2.4 初步确定在胃癌中具有重要意义的 circRNA

运用生物信息学软件 Targetscan 和 Miranda 对 circRNA 与 miRNA 相互作用靶点进行预测，每条 circRNA 均预测到 5 个 miRNA 结合位点（表 4、5、图 3）。在所有筛选出来的 circRNA 中，我们通过文献检索发现，hsa_circ_0001897 已被报导与胃癌分期、转移相关，与其结合的 miR-150-5p 已被证实与多种消化道肿瘤密切相关，如结肠癌、肝癌、胆

表1. 总RNA与cRNA的质控

Table 1. Quality assurance of total RNA and cRNA quantification

Sample	Total RNA					cRNA			
	OD ₂₆₀ /OD ₂₈₀	OD ₂₆₀ /OD ₂₃₀	Quantity (ng)	Volume (μL)	Concentration (ng/μL)	Cy3 positive (pmol Cy3/μg)	Quantity (μg)	Volume (μL)	Concentration (μg/μL)
GES-1.1	1.92	2.47	78 352.80	120	652.94	31.283 53	4.286 6	10	0.428 66
GES-1.2	1.94	2.46	56 376.80	80	704.71	29.639 95	4.666 0	10	0.466 60
GES-1.3	1.94	2.44	67 057.60	80	838.22	27.396 94	4.318 0	10	0.431 80
MGC-803.1	1.95	2.45	117 630.00	120	980.25	31.554 98	4.611 0	10	0.461 10
MGC-803.2	1.95	2.45	88 435.20	120	736.96	30.708 55	4.842 3	10	0.484 23
MGC-803.3	1.98	2.41	142 429.20	120	1 186.91	29.985 41	4.455 5	10	0.445 55
SGC-7901.1	1.97	2.38	111 660.00	120	930.50	30.671 96	4.460 1	10	0.446 01
SGC-7901.2	1.96	2.42	91 425.60	120	761.88	30.336 36	4.545 7	10	0.454 57
SGC-7901.3	1.96	2.44	151 070.40	160	944.19	28.344 02	4.007 9	10	0.400 79
NCI-N87.1	1.97	2.49	129 976.00	160	812.35	30.792 14	4.968 8	10	0.496 68
NCI-N87.2	1.98	2.42	152 438.40	160	952.74	29.049 60	4.093 0	10	0.409 30
NCI-N87.3	2.00	2.37	146 052.80	160	912.83	29.659 04	4.507 9	10	0.450 79

For spectrophotometer, the OD₂₆₀/OD₂₈₀ ratio should be close to 2.0 for pure RNA (ratios between 1.8 and 2.1 are acceptable). The OD₂₆₀/OD₂₃₀ ratio should be more than 1.8. If the yield is < 1.65 μg and the specific activity is < 9.0 pmol Cy3/μg, cRNA do not proceed to the hybridization step.

表2. 胃癌细胞和正常胃上皮细胞中差异表达上调的前5条circRNA

Table 2. Top 5 up-regulated expressed circRNAs between gastric cancer cells and normal gastric epithelial cells

No.	circRNA	Fold change	P	circRNA type
1	hsa_circ_0003057	2.690 924 5	0.000 013 390 24	Exonic
2	hsa_circ_0005044	2.690 082 6	0.008 701 295 95	Intronic
3	hsa_circ_0001944	2.654 759 2	0.003 374 118 79	Exonic
4	hsa_circ_0091537	2.495 864 3	0.000 877 223 01	Exonic
5	hsa_circ_0001829	2.288 660 7	0.032 868 960 87	Exonic

circRNA type: The location of the circRNA in the chromosome.

表3. 胃癌细胞和正常胃上皮细胞差异表达下调的前5条circRNA

Table 3. Top 5 down-regulated expressed circRNAs between gastric cancer cells and normal gastric epithelial cells

No.	circRNA	Fold change	P	circRNA type
1	hsa_circ_0022382	12.279 568 4	0.000 273 211 7	Exonic
2	hsa_circ_0000994	10.698 419 1	0.000 002 434 35	Exonic
3	hsa_circ_0022383	10.386 687 1	0.000 454 694 93	Exonic
4	hsa_circ_0007444	8.968 459 1	0.002 682 606 23	Exonic
5	hsa_circ_0005232	8.295 784	0.000 001 892 63	Exonic

circRNA type: The location of the circRNA in the chromosome.

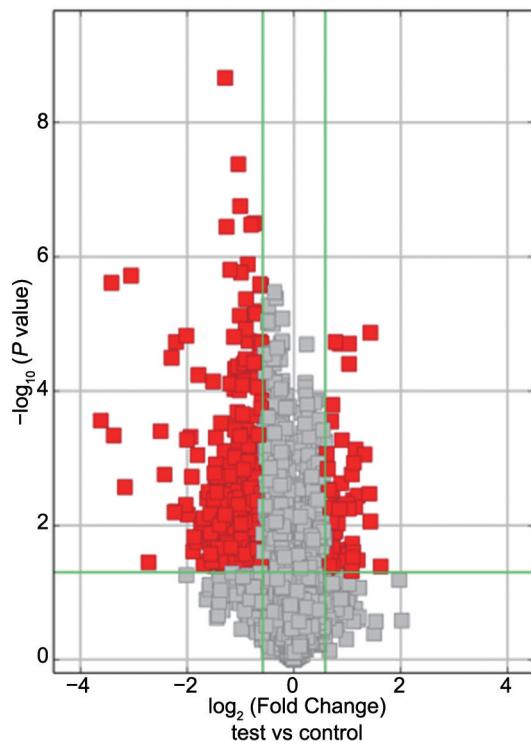


图 1. circRNA火山图

Fig. 1. Volcano plot of circRNAs. The red points in plot represent the differentially expressed circRNAs with statistical significance between gastric cancer cells and normal gastric epithelial cells. The left red points represent down-regulation, and the right red points represent up-regulation.

管瘤^[6-8];与hsa_circ_0008106、hsa_circ_0060456存在结合位点的miR-16-5p已被证实参与胃癌的发生和发展，并且与肿瘤恶性程度相关^[9,10]。

3 讨论

CircRNA作为非编码RNA家族的新成员，正吸引着越来越多人的注意。生物信息技术的飞速发展将人们带入circRNA的世界，人们对circRNA的认识正在不断深入，其许多生物学特性已被发现，其中一些重要的特征包括广泛性、高度稳定性及保守性等^[11-14]。目前，研究已经证实circRNA的差异表达与包括胃癌在内的恶性肿瘤密切相关。CircRNA广泛性、高度稳定性等生物学特征及在肿瘤中的表达特点提示我们：它可能作为新型肿瘤的生物标志物并对肿瘤的诊断起到支持作用。CircRNA在肿瘤中的分子作用机制有待进一步研究。

最近，有学者提出竞争性内源RNA(competting endogenous RNA, ceRNA)假说，认为具有相同miRNA应答元件(miRNA response element, MRE)的RNA

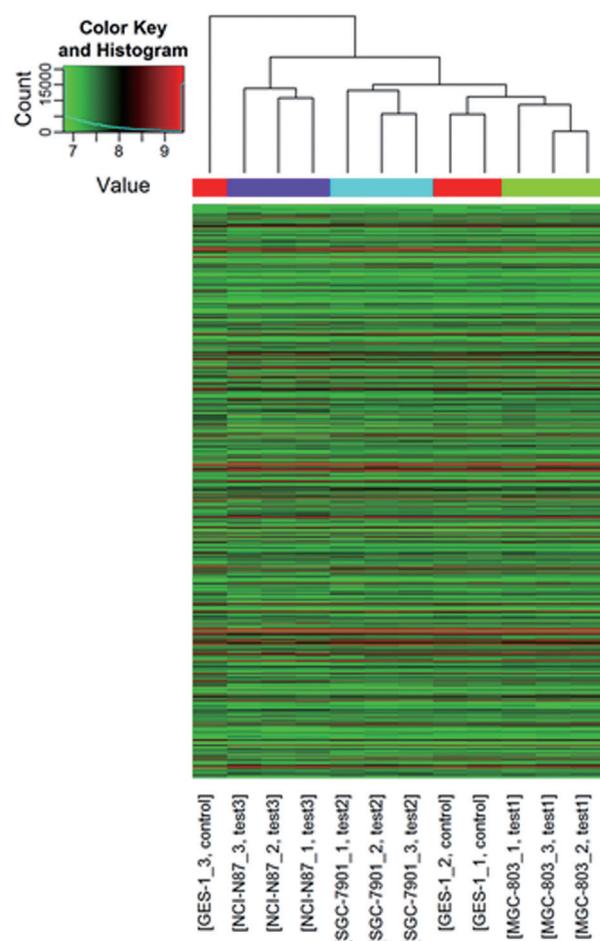


图 2. 差异circRNA聚类图

Fig. 2. Clustering plot of the differentially expressed circRNAs between gastric cancer cells and normal gastric epithelial cells. The result from unsupervised hierarchical clustering analysis shows distinguishable circRNA expression profiling among samples. Each column represents the expression profile of a cell sample, and each row corresponds to a circRNA. “Red” indicates higher expression level, and “green” indicates lower expression level.

分子，可以通过竞争性地结合相同的miRNA，实现对miRNA的靶向调控^[15]。CircRNA作为ceRNA家族新发现的重要成员，本身富含miRNA结合位点，可发挥“海绵作用”吸附miRNA，降低miRNA对靶基因的抑制作用，增加靶基因的表达水平，进而起到调控生物学过程的作用^[11,16]。Cao等在研究肝癌时观察到，circMTO1能够靶向结合miR-9来调控p21的表达，从而抑制肿瘤细胞的增殖和侵袭^[17]。Chen等在研究口腔癌时观察到，circRNA_100290可通过靶向结合miR-29，从而解除miR-29对CDK6的抑制，促进口腔鳞状细胞癌的发生和发展^[18]。

表4. hsa_circ_0001897、hsa_circ_0008106和hsa_circ_0060456的详细信息

Table 4. Details of hsa_circ_0001897, hsa_circ_0008106 and hsa_circ_0060456

circRNA	Fold change (down-regulated)	P	circRNA type	miRNA binding site
hsa_circ_0001897	1.846 206 2	0.000 065 301 39	Exonic	hsa-miR-150-5p hsa-miR-29a-3p hsa-miR-29c-3p hsa-miR-29b-3p hsa-miR-767-5p
hsa_circ_0008106	2.055 149 4	0.001 133 053 95	Exonic	hsa-miR-29b-2-5p hsa-miR-29a-5p hsa-miR-518c-5p hsa-miR-214-3p hsa-miR-16-5p
hsa_circ_0060456	1.861 731 1	0.001 182 505 61	Exonic	hsa-miR-647 hsa-miR-412-3p hsa-miR-637 hsa-miR-660-3p hsa-miR-16-5p

circRNA type: The location of the circRNA in the chromosome. For each circRNA, we identified 5 miRNAs with the highest mirSVR score to establish a “Top-5” circRNA-miRNA network (1 circRNA connecting to 5 miRNAs)

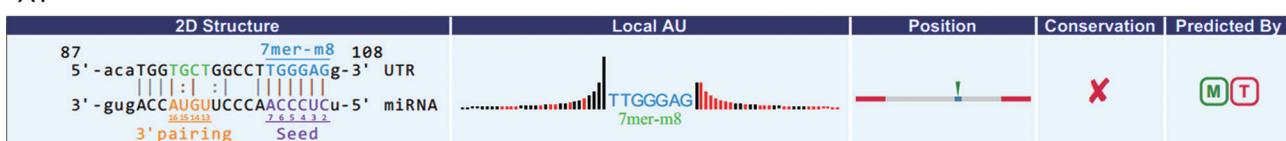
表5. hsa_circ_0001897、hsa_circ_0008106和hsa_circ_0060456在胃癌细胞中的表达

Table 5. The expression of hsa_circ_0001897, hsa_circ_0008106 and hsa_circ_0060456 in gastric cancer cells

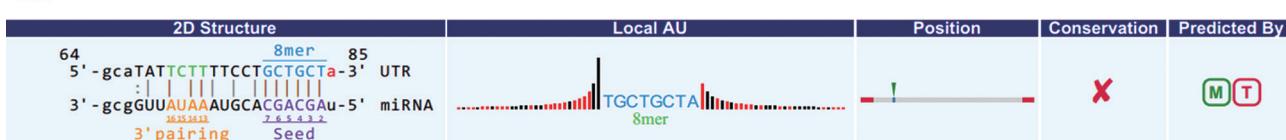
circRNA	Fold change		
	MGC-803 vs GES-1	SGC-7901 vs GES-1	NCI-N87 vs GES-1
hsa_circ_0001897	1.950 956 1	1.756 716 1	1.836 079 7
hsa_circ_0008106	1.834 483 0	2.222 134 8	2.129 345 3
hsa_circ_0060456	1.505 752 4	1.804 972 4	2.374 251 7

GES-1 cells were normal gastric epithelial cells and used as the control group.

A1



A2



A3

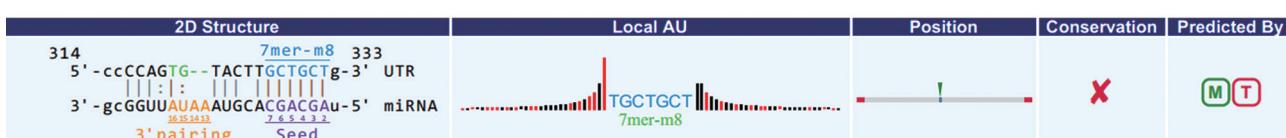


图 3. circRNA与miRNA相互作用位点的详细注释图

Fig. 3. The detailed annotation for circRNA-miRNA interactions. A1: Complementary situations of hsa_circ_0001897 and miR-150-5p. A2: Complementary situations of hsa_circ_0008106 and miR-16-5p. A3: Complementary situations of hsa_circ_0060456 and miR-16-5p.

在胃癌相关 circRNA 的研究方面, Zhu 等利用高通量 RNA 测序技术观察到, 与癌旁组织对比, circLARP4 在胃癌组织中明显低表达; 通过生物信息学技术观察到 circLARP4 与 miR-424-5p 存在结合位点, LATS1 为其下游靶基因, 随后借助双荧光素酶实验和 RNA 免疫共沉淀技术验证三者之间的结合位点, 最后通过 MTT、克隆增殖等实验观察到 circLARP4 通过结合 miR-424-5p 来调控 LATS1 的表达, 进而抑制胃癌细胞的增殖与侵袭^[19]。Li 等利用 circRNA 基因芯片技术观察到 46 条表达差异显著的 circRNA, 随后选定低表达的 circRNA-100269 为研究对象, 利用 RT-PCR 验证了芯片结果, 随后通过生物信息学软件预测 circRNA-100269 与 miR-630 之间可能存在结合位点, 在后续的分子生物学功能实验中, 通过表达实验观察到 circRNA-100269 能够调控 miR-630 的表达, 从而抑制胃癌细胞的增殖, 并且发现 miR-630 高表达能够阻止 circRNA-100269 对胃癌细胞的抑制效应^[20]。Guo 等亦认为 hsa_circ_0000190 可能作为新的胃癌诊断和治疗标志物, 相比传统的胃癌肿瘤标志物 CEA、CA19-9, hsa_circ_0000190 敏感性和特异性均较高^[21]。

本研究借助 Arraystar 公司 circRNA 芯片筛选不同分化程度胃癌细胞 MGC-803、SGC-7901、NCI-N87 与正常胃上皮细胞 GES-1 间差异表达的 circRNA。结果显示, 与正常胃上皮细胞相比, 在三种不同分化程度的胃癌细胞中均表达上调的 circRNA 有 79 条, 均表达下调的有 229 条。在这些差异表达的 circRNA 中, 我们通过文献检索并挑选了具有研究价值的 circRNA: hsa_circ_0001897 在胃癌细胞中表达下调, 已有文献报道其与胃癌分期、转移相关^[22], 但它在胃癌中的分子作用机制还未被阐明, 通过生物信息学软件 Targetscan 和 Miranda 预测, hsa_circ_0001897 与 miR-150-5p 存在结合位点, 进一步检索发现, miR-150-5p 与多种消化道肿瘤密切相关, 如结肠癌、肝癌、胆管癌^[6-8], 由此, 我们推测 hsa_circ_0001897 可能通过调控 miR-150-5p 抑制胃癌细胞的侵袭与转移; hsa_circ_0008106、hsa_circ_006045 在胃癌细胞中表达下调, 与其存在结合位点的 miR-16-5p 已被证实参与胃癌的发生和发展^[9,10], 这些结果提示我们, hsa_circ_0008106、hsa_circ_0060456 可能通过与 miR-16-5p 相互作用, 从而抑制胃癌细胞的侵袭与转移, 目前, hsa_circ_0008106、hsa_circ_0060456 还未曾被报道。后

续的研究应深入探讨 hsa_circ_0001897、hsa_circ_0008106 和 hsa_circ_0060456 在胃癌中的分子作用机制, 以期为寻找胃癌新的诊治方法和药物治疗靶点提供理论基础。

参考文献

- 1 Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer* 2015; 136(5): E359–E386.
- 2 Ashwal-Fluss R, Meyer M, Pamudurti NR, Ivanov A, Bartok O, Hanan M, Evental N, Memczak S, Rajewsky N, Kadener S. circRNA biogenesis competes with pre-mRNA splicing. *Mol Cell* 2014; 56(1): 55–66.
- 3 Weng W, Wei Q, Toden S, Yoshida K, Nagasaka T, Fujiwara T, Cai S, Qin H, Ma Y, Goel A. Circular RNA ciRS-7—a promising prognostic biomarker and a potential therapeutic target in colorectal cancer. *Clin Cancer Res* 2017; 23(14): 3918–3928.
- 4 Qin M, Liu G, Huo X, Tao X, Sun X, Ge Z, Yang J, Fan J, Liu L, Qin W. Hsa_circ_0001649: a circular RNA and potential novel biomarker for hepatocellular carcinoma. *Cancer Biomark* 2016; 16(1): 161–169.
- 5 Yao JT, Zhao SH, Liu QP, Lv MQ, Zhou DX, Liao ZJ, Nan KJ. Over-expression of CircRNA_100876 in non-small cell lung cancer and its prognostic value. *Pathol Res Pract* 2017; 213(5): 453–456.
- 6 Wang WH, Chen J, Zhao F, Zhang BR, Yu HS, Jin HY, Dai JH. MiR-150-5p suppresses colorectal cancer cell migration and invasion through targeting MUC4. *Asian Pac J Cancer Prev* 2014; 15(15): 6269–6273.
- 7 Li T, Xie J, Shen C, Cheng D, Shi Y, Wu Z, Zhan Q, Deng X, Chen H, Shen B, Peng C, Li H, Zhu Z. miR-150-5p inhibits hepatoma cell migration and invasion by targeting MMP14. *PLoS One* 2014; 9(12): e115577.
- 8 Wu X, Xia M, Chen D, Wu F, Lv Z, Zhan Q, Jiao Y, Wang W, Chen G, An F. Profiling of downregulated blood-circulating miR-150-5p as a novel tumor marker for cholangiocarcinoma. *Tumour Biol* 2016; 37(11): 15019–15029.
- 9 Zhang J, Song Y, Zhang C, Zhi X, Fu H, Ma Y, Chen Y, Pan F, Wang K, Ni J, Jin W, He X, Su H, Cui D. Circulating MiR-16-5p and MiR-19b-3p as two novel potential biomarkers to indicate progression of gastric cancer. *Theranostics* 2015; 5(7): 733–745.
- 10 Zhu C, Huang Q, Zhu H. Melatonin inhibits the proliferation of gastric cancer cells through regulating the miR-16-5p-Smad3 pathway. *DNA Cell Biol* 2018; 37(3): 244–252.

- 11 Long Z (龙志), Xie J, Li GQ. Research progress of circular RNA in human diseases. *J Milit Surg Southwest Chin (西南军医)* 2018; 20(1): 40–44 (in Chinese).
- 12 Jeck WR, Sorrentino JA, Wang K, Slevin MK, Burd CE, Liu J, Marzluff WF, Sharpless NE. Circular RNAs are abundant, conserved, and associated with ALU repeats. *RNA* 2013; 19(2): 141–157.
- 13 Rybak-Wolf A, Stottmeister C, Glazar P, Jens M, Pino N, Giusti S, Hanan M, Behm M, Bartok O, Ashwal-Fluss R, Herzog M, Schreyer L, Papavasileiou P, Ivanov A, Ohman M, Refojo D, Kadener S, Rajewsky N. Circular RNAs in the mammalian brain are highly abundant, conserved, and dynamically expressed. *Mol Cell* 2015; 58(5): 870–885.
- 14 Chen LL. The biogenesis and emerging roles of circular RNAs. *Nat Rev Mol Cell Biol* 2016; 17(4): 205–211.
- 15 Salmena L, Poliseno L, Tay Y, Kats L, Pandolfi PP. A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language? *Cell* 2011; 146(3): 353–358.
- 16 Ebert MS, Neilson JR, Sharp PA. MicroRNA sponges: competitive inhibitors of small RNAs in mammalian cells. *Nat Methods* 2007; 4(9): 721–726.
- 17 Han D, Li J, Wang H, Su X, Hou J, Gu Y, Qian C, Lin Y, Liu X, Huang M, Li N, Zhou W, Yu Y, Cao X. Circular RNA circMTO1 acts as the sponge of microRNA-9 to suppress hepatocellular carcinoma progression. *Hepatology* 2017; 66(4): 1151–1164.
- 18 Chen L, Zhang S, Wu J, Cui J, Zhong L, Zeng L, Ge S. circRNA_100290 plays a role in oral cancer by functioning as a sponge of the miR-29 family. *Oncogene* 2017; 36(32): 4551–4561.
- 19 Zhang J, Liu H, Hou L, Wang G, Zhang R, Huang Y, Chen X, Zhu J. Circular RNA_LARP4 inhibits cell proliferation and invasion of gastric cancer by sponging miR-424-5p and regulating LATS1 expression. *Mol Cancer* 2017; 16(1): 151.
- 20 Zhang Y, Li J, Yu J, Liu H, Shen Z, Ye G, Mou T, Qi X, Li G. CircRNA_100269 is downregulated in gastric cancer and suppresses tumor cell growth by targeting miR-630. *Aging (Albany NY)* 2017; 9(6): 1585–1594.
- 21 Chen S, Li T, Zhao Q, Xiao B, Guo J. Using circular RNA hsa_circ_0000190 as a new biomarker in the diagnosis of gastric cancer. *Clin Chim Acta* 2017; 466: 167–171.
- 22 Shao Y, Li J, Lu R, Li T, Yang Y, Xiao B, Guo J. Global circular RNA expression profile of human gastric cancer and its clinical significance. *Cancer Med* 2017; 6(6): 1173–1180.