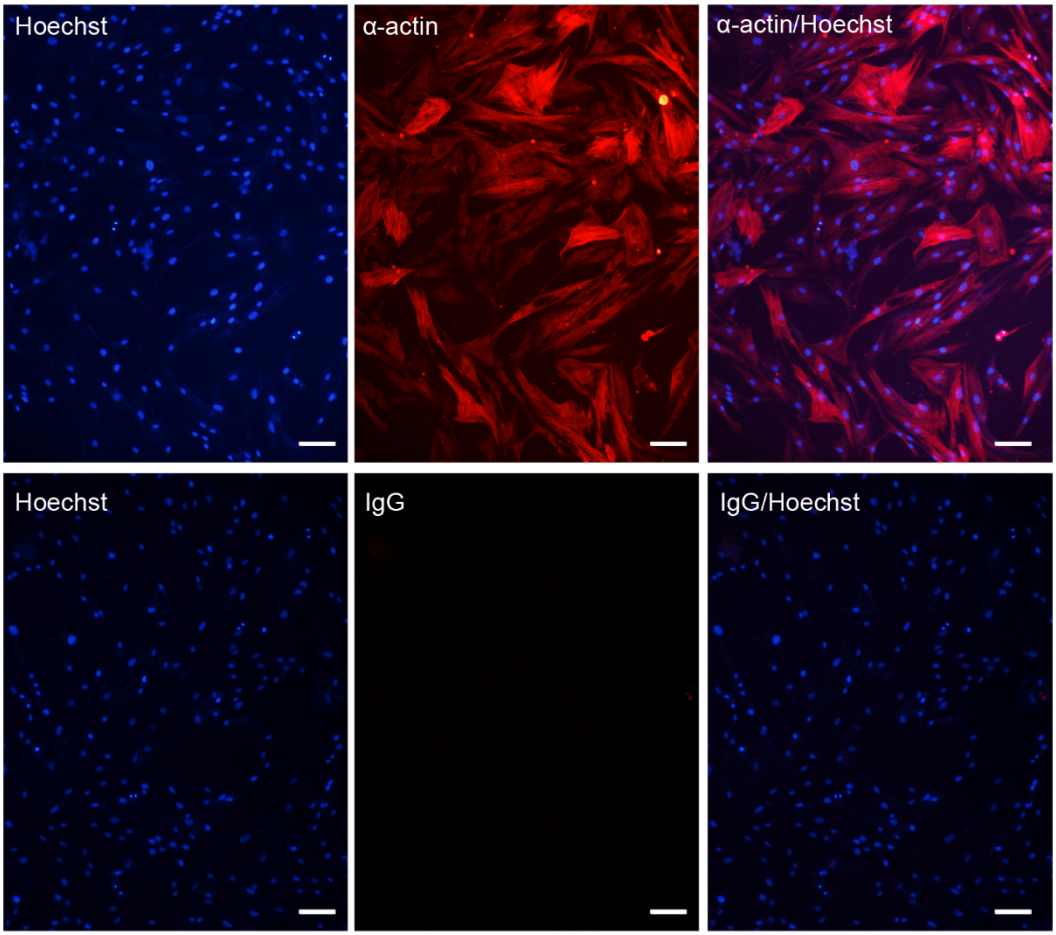
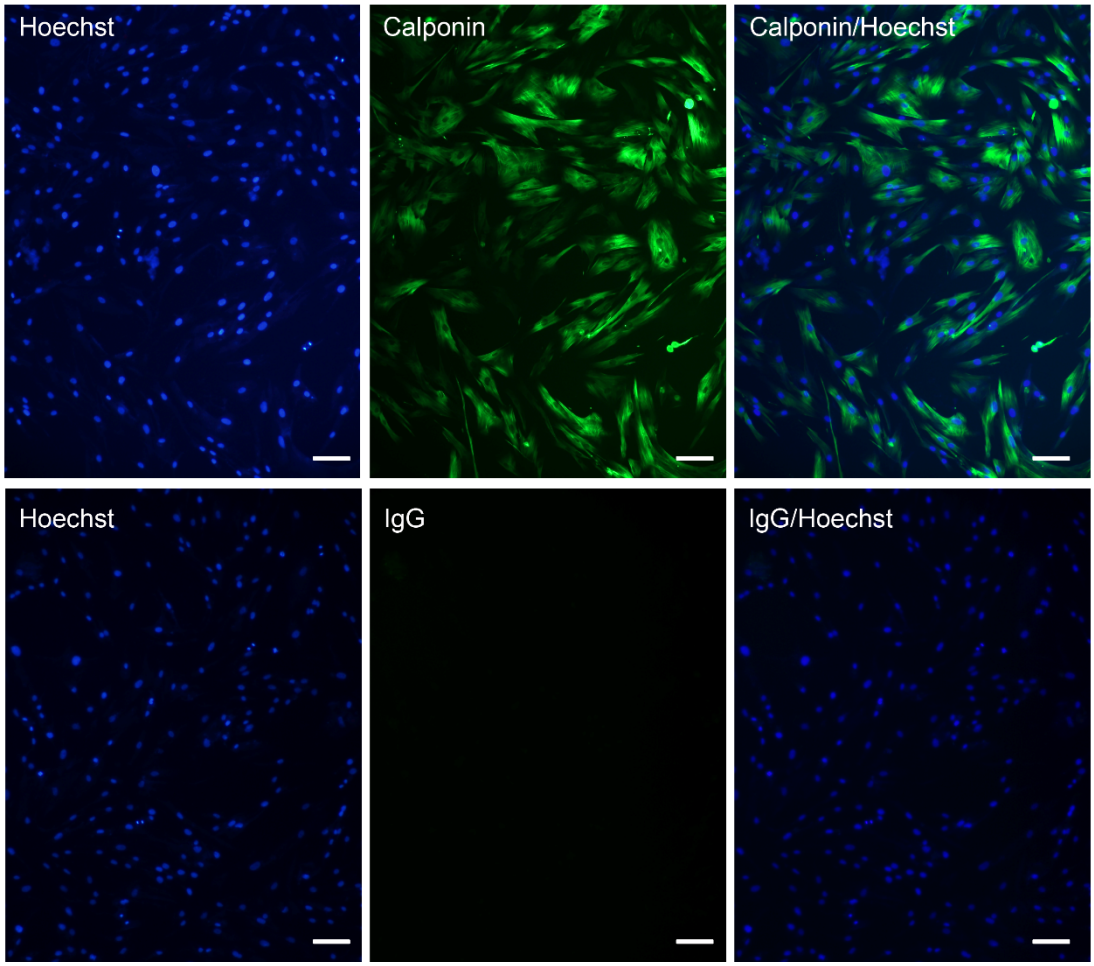


Supplement Fig. 1. Immunofluorescence staining for α-smooth muscle actin in cultured HUSMCs. Cells were fixed in 4% paraformaldehyde solution for 20 min at room temperature. The fixed cultures were washed with PBS and incubated with 10% BSA for 2 h and then were incubated with monoclonal antibody recognizing α-smooth muscle actin (α-actin, sc-58669, Santa Cruz) at the dilution of 1:100 in PBS containing 2% BSA overnight at 4 °C. Subsequently, the specimens were incubated with anti-mouse IgG conjugated to Alexa Fluor 594 (1:400 dilution, Invitrogen). The cell nuclei were visualized by applying the DNA-specific dye Bisbenzimide Hoechst 33342 (1:5 000, Sigma-Aldrich). For negative controls, the primary antibody was substituted with a normal IgG in the same dilution. Staining images were visualized using the Olympus fluorescent microscope (Tokyo, Japan). Scale bar, 100 μm.



Supplement Fig. 2. Immunofluorescence staining for calponin in cultured HUSMCs. Cells were fixed in 4% paraformaldehyde solution for 20 min at room temperature. The fixed cultures were washed with PBS and incubated with 10% BSA for 2 h and then were incubated with monoclonal antibody recognizing calponin (Abcam, ab46794) at the dilution of 1:100 in PBS containing 2% BSA overnight at 4 °C. Subsequently, the specimens were incubated with anti-rabbit IgG conjugated to Alexa Fluor 488 (1:400 dilution, Invitrogen). The cell nuclei were visualized by applying the DNA-specific dye Bisbenzimide Hoechst 33342 (1:5 000 dilution, B2261, Sigma-Aldrich). For negative controls, the primary antibody was substituted with a normal IgG in the same dilution. Staining images were visualized using the Olympus fluorescent microscope (Tokyo, Japan). Scale bar, 100 μm.